

REVIEW: MOLECULAR CELL BIOLOGY

DECODING CALCIUM SIGNALS IN LIVING CELLS

Cedric Viero^{1*} and Govindan Dayanithi²

¹Department of Cardiology, Wales Heart Research Institute, School of Medicine, Cardiff University, Cardiff, UK ²Department of Cellular Neurophysiology, Institute of Experimental Medicine, Academy of Sciences of the Czech Republic, European Union Centre of Excellence, Prague, CZ

Received on: 26th-April-2010; Revised on: 2nd-May-2010; Accepted on: 2nd-May-2010; Published on: 2nd-May-2010. *Corresponding author: Email: VieroCL@cardiff.ac.uk Tel: +44-2920744046 ; Fax: +44-2920743500

ABSTRACT

The regulation of calcium ion concentration outside and inside the cell, together with the movement of calcium ions from one compartment to another, are key factors that determine the fate of a cell: growth, stimulation and death. Calcium is the most ubiquitous second messenger encountered in a plethora of physiological functions from hormonal release and muscular contraction to gene expression. The level of increase in calcium concentration within a cell is finely modulated in terms of the quantity of ions, time and space, thus constituting a complex language that can only be interpreted by specific proteins.

Keywords: Signalling; Excitation-contraction coupling; Exocytosis; Microdomains; Ion channels

[I] PHYSIOLOGICAL FUNCTIONS

The homeostasis of calcium ion concentration within a cell is a fine-tuned regulated parameter which presides over many pivotal physiological functions. For example, in the heart, the contraction of cardiac muscle cells is triggered by depolarizing events that induce a small influx of calcium ions, which (through L-type calcium channels or LCCs) in turn leads to the massive release of calcium from intracellular pools (sarcoplasmic reticulum or SR) via calcium release channels known as ryanodine receptors (RyRs). This released calcium binds to the contractile apparatus, which mediates a conformational change and the movement of the myofilaments and therefore a shortening of the cardiomyocyte. This process is referred to as excitation-contraction coupling [1]. In contrast, skeletal muscle cells exhibit a direct mechanical coupling between LCCs and RyRs [2]. In neurones, the cellular localization of calcium channels determines their particular function. At presynaptic sites, calcium channels regulate the release of neurotransmitters by exocytosis. In dendritic spines (that contain the post-synaptic densities where the neurotransmitter receptor complexes are present), calcium signals are responsible for the modulation of learning and memory processes [3]. In particular intracellular inositoltrisphosphate receptors (or InsP₃Rs, calcium release channels that are located on the membrane of the endoplasmic reticulum or ER) play a major role as signal detectors and integration centres [4]. In non-excitable cells, $InsP_3Rs$ are the central components of the calcium-induced calcium release (CICR)

phenomenon, but not the only components, since RyRs can also be present and functional such as in the pancreas where RyRs mediate calcium release [5].

Calcium signals can be found in various compartments and organelles of the cell where they display distinct functions. Mitochondria, for instance, serve as calcium buffering systems, and alteration of this pathway leads to pathophysiological states in many varieties of tissues such as in the heart, brain, pancreas and kidney [6]. Moreover, lysosomes also represent calcium stores, and the calcium release from acidic organelles is mediated by nicotinic acid adenine dinucleotide phosphate (NAADP) receptors identified as two-pore channels [7]. It is now accepted that the nucleus itself is involved in the occurrence of calcium signals [8, 9]. These nucleoplasmic calcium signals activate distinct pathways that control gene expression very specifically and independently of cytosolic calcium [10].

Hence, calcium controls many aspects of life at the cellular but also at the organism level, from fertilization of mammalian, sea urchin, fish and frog eggs through development, differentiation and proliferation to the activation of transcription factors and apoptosis [11].

[II] PATHWAYS INVOLVED IN THE INCREASE OF THE INTRACELLULAR CALCIUM CONCENTRATION

There are 3 main pathways by which a rise in intracellular calcium can occur. They are: i) calcium entry through voltage-gated calcium channels, ii) calcium entry through ligand-gated channels, and iii) calcium release from internal stores.

While the voltage-gated, ligand-gated and store depletion pathways have been studied intensively, the involvement of polymodal (responsive to temperature, light, voltage, ligand, pH and mechanical stimuli) transient receptor potential (or TRP) channels has been the subject of recent investigations [12-14]. These channels are thought to be involved in many physiological functions such as blood pressure, the regulation of mineral absorption/reabsorption, gut motility and airway responsiveness, pain and taste transductions, thermo- and mechano-sensations and cell proliferation/death [15].

In many neuronal and non-neuronal cell types, various molecules (such as peptides, hormones and transmitters) have been shown to specifically bind to their receptors and upon stimulation trigger an influx of calcium *via* activation of phospholipase C (PLC) and adenylate cyclase (AC) pathways, thus exerting cellular functions [16]. Calcium release from stores in mammalian cell types primarily occurs as a result of the activation of metabotropic ligand-binding receptors, which use second messenger signalling cascades to indirectly activate intracellular ion channels.

[III] DIFFERENT TYPES OF CALCIUM SIGNALS

3.1. Calcium sparklets

Optical recordings of localized calcium influx events *via* LCC present in the plasma membrane are called "calcium sparklets". When a single LCC opens, this leads to calcium influx, resulting in an increase in the total amount of calcium in the cleft between the LCC in the plasma membrane and the opposing cluster of RyRs in the SR membrane. These increases can be recorded as calcium signals named calcium sparklets [17]. These sparklets reveal an unexpected feature of these channels, depending on their activity. In cerebral arterial myocytes, they are of two types, i.e. exhibiting low or high persistent activity, and both are activated by PKC/PKA [18]. Highly elevated PKA-dependent calcium sparklets are observed in cerebral arterial smooth muscle during acute hyperglycemia, vascular dysfunction and diabetes.

3.2. Calcium sparks

The cluster of RyRs that faces the sparklet senses the calcium ions, leading to the synchronous opening of the entire cluster of RyRs and to a highly localised release of calcium out of the SR into the membrane cleft [19]. These elementary signals are thought to sum up to underlie global calcium transients to initiate contraction in the heart. Calcium sparks are usually characterised by the amplitude, the duration, the rise time, the spatial spread of the Ca^{2+} signal, and by the frequency (of



occurrence of events) of individual spark sites or the frequency of all the sparks within one cell [20]. In smooth muscles, sparks can occur spontaneously or after activation by many factors such as caffeine, calcium entry via L-type calcium channels, increase in SR calcium content and stretch (see for details: [21]). It should be noted that calcium sparks are the only small physiologically relevant elementary calcium release events in cellular calcium signalling and that the corresponding homogeneous calcium release remains unexplained. However, calcium sparks have been shown to be a major signalling pathway for excitation-contraction coupling.

3.3. Calcium quarks

There is yet another phenomenon in which highly localized SR calcium release in both skeletal and cardiac muscles may be triggered under certain conditions, i.e. calcium quarks [22]. By using two-photon photolytic activation of localized calcium release in cardiac myocytes, it has been shown that a single calcium spark actually consists of the summation of unitary calcium events characterised by tiny amplitudes, a very short lifetime and tight spatial confinement. Hence, each individual RyR could give rise to calcium releases in the dyadic cleft called calcium quarks [22]. However, this phenomenon remains unclear (see also: [23]).

3.4. Calcium puffs

Local intracellular calcium signals caused by the opening of clustered InsP3 receptors in the ER or SR membrane are known as calcium puffs [8, 24]. Calcium puffs display a longer lifetime in comparison to calcium sparks and a wider spread. Each calcium puff would be composed of elementary events named calcium blips [25]. Calcium puff events have been characterised in neurones [26] and in smooth muscle cells [27].

3.5. Calcium marks, scraps and blinks

Microdomain miniature calcium transients in single mitochondria are termed "marks" and SR luminal calcium depletion transients are termed "scraps".

In the cardiac cell line H9C2, it has been shown that subsequent to calcium spark events, rapid increases in calcium in the neighbouring mitochondria can occur [28]. It was proposed that calcium marks are triggered by RyRs through a process involving the travel of calcium from spark sites into the mitochondria.

Attempts were made to monitor calcium movements within the SR lumen, which led to the observation of local depletions of the intracellular calcium concentration ("scraps"), mirrors of the calcium transients recorded in the cytosol [29]. Likewise, rapid and substantial depletions of calcium from the nanometer-sized luminal stores in heart cells have been shown and called "calcium blinks", blinks meaning the confined SR depletions from single calcium sparks [30, 31]. Therefore, the visualization of these local store calcium signals can be an important key to understand cardiac physiopathology.

3.6. Calcium waves

Calcium waves are global increases of the intracellular calcium concentration from the SR or ER that propagate throughout the cell, sustained by the phenomenon of CICR. These events have a longer lifetime and a larger amplitude than the elementary events previously described and can appear in several forms (spirals, U- and V-shapes and circular waves). The analysis of their kinetics gives indications about the activity of proteins involved in buffering systems such as SR/ER calcium ATPase (SERCA) pumps [32].

All these different types of calcium signals modulate various physiological functions specific to particular cell types.

[IV] CALCIUM HANDLING MECHANISMS

The common method to detect changes in the intracellular calcium concentration is to use fluorescent calcium probes (Fura-2, Fluo-3, Fluo-4, Indo-1, Calcium Green, Rhod-2, or Oregon Green 488 BAPTA-1, this list being obviously not exhaustive) and to monitor the corresponding signals over time. However, one has to bear in mind that such techniques only enable the investigator to detect free calcium ions, i.e. calcium which is not bound to proteins, whereas most of the calcium ions in the cell are bound. The role of such calciumbinding proteins is not only important for creating a buffering system, but also for interpreting calcium signals and triggering molecular responses accordingly. Indeed, it seems difficult to relate the individual, short and sometimes highly localized increases of calcium concentration to other signalling processes such as phosphorylation events, which one assumes would be more global and of longer onset/duration.

Recently, a nice study has provided the beginning of an answer to that question. The authors expressed the calcium-dependent protein kinase C (PKC) isoform alpha fused to various fluorescent proteins in HEK293 and COS1 cells. By confocal imaging, they could monitor the PKC translocation events from the cytosol to the plasma membrane. They successfully demonstrated for the first time the relationship between global and local calcium signals on the one hand, and global and local PKC translocation events on the other hand. It should be noted that they evidenced that calcium signals and PKC translocations were concomitant [33].



Therefore, PKCalpha is a cellular "calcium sensor" following the spatio-temporal characteristics of each calcium signal. Whether these PKC translocation events are associated with phosphorylation events is extremely likely but still needs to be investigated.

Calcium fluctuations are sensed by a wide array of "calcium sensors" which have one common feature: they are all endowed with calcium-binding sites, such as in calciumsensitive enzymes, with different affinities for calcium, and the binding of the latter modifies their activity by changing their conformation, which therefore triggers or discontinues diverse biochemical processes, which in turn control various cellular reactions [34]. The duration and kinetics of intracellular calcium fluctuations control the timing of calciumbinding/unbinding to the "calcium sensors", thus forming the basis for the temporal coding of calcium signalling events [35]. There are a few main intracellular compartments intimately involved in creating, encoding and decoding calcium signals, which are the cytoplasm, the Golgi complex, lysosomes, secretory granules, the nucleus, the endoplasmic reticulum and the mitochondria [36]. Furthermore, the formation of intracellular free calcium fluctuations is controlled by calcium channels and transporters, which, being activated by physiological stimulations, precisely regulate the calcium ion concentration [35].

[V] CALCIUM CLEARANCE MECHANISMS

Calcium clearance mechanisms are important to maintain calcium homeostasis within a range of concentrations that allow biochemical reactions to take place but are not toxic to cellular functions [37]. Indeed calcium removal (or calcium clearance) is even decisive for the fate of the cell, which includes the "cell death" process when the cell has lost the capacity to extrude or reuptake the extra amount of calcium [38, 39]. The mechanisms involved in intracellular calcium buffering and restoration to basal levels include calcium pumps in both intracellular compartments and the plasma membrane, mitochondrial calcium uptake, plasma membrane sodium/calcium exchange and calcium-binding proteins in the cytosol [Figure-1]. Depending on the cell type, the mechanisms of calcium homeostasis and the implications of each of the clearance systems mentioned above will be different.



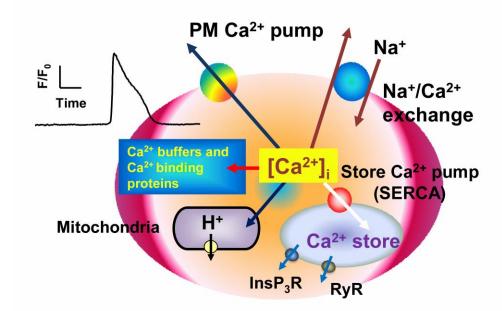


Fig: 1. Schematic representation of possible calcium clearance mechanisms in neuronal, muscle and non-excitable cells: Arrows suggest that increased cytosolic calcium can be extruded *via* plasma membrane (PM) Ca^{2+} pump, Na^+/Ca^{2+} exchange, stored by mitochondria, intracellular compartments (*via* sarco/endoplasmic reticulum Ca^{2+} pump, SERCA), Ca^{2+} buffers and Ca^{2+} binding proteins. Abbreviations: ryanodine receptor (RyR), inositol-trisphosphate receptor (InsP₃R), intracellular calcium concentration ([Ca^{2+}]_i) and normalized change of the calcium-dependent fluorescence (F/F₀). A representative [Ca^{2+}]_i transient (recorded with a fluorescent calcium indicator) is depicted in the top left corner of the cartoon.

[VI] CONCLUSION

Calcium is the key to a wide variety of cellular processes. Monitoring and deciphering calcium signals can be a useful tool to predict how cells are likely to behave. Besides the understanding of essential physiological phenomena, the exploration of the readout of calcium events by calcium detectors (fluorescent or endogenous calcium-binding proteins) could have applications in detecting pathophysiological remodelling, arrhythmic cardiac patterns and neurological disorders such as Alzheimer's disease [40]. Once typical calcium signal patterns can be identified and defined, this will be useful in developing drug screening strategies and drug therapies.

FINANCIAL DISCLOSURE

The authors state that they have no conflict of interest pertaining to this manuscript.

ACKNOWLEDGEMENT

We are grateful to James Dutt (IEM, Prague, Czech Republic) for critical reading and language editing of the manuscript.

REFERENCES

- [1] Bers D. [2002] Cardiac excitation-contraction coupling. *Nature* 415(6868):198-205.
- [2] Paolini C, Quarta M, Nori A, Boncompagni S, Canato M, Volpe P, et al. [2007] Reorganized stores and impaired calcium handling in skeletal muscle of mice lacking calsequestrin-1. *J Physiol* 583(Pt 2):767-84.
- [3] Berridge M. [1998] Neuronal calcium signaling. *Neuron* 21(1):13-26.
- [4] Nakamura T, Barbara J, Nakamura K, Ross W. [1999] Synergistic release of Ca2+ from IP3-sensitive stores evoked by synaptic activation of mGluRs paired with backpropagating action potentials. *Neuron* 24(3):727-37.
- [5] Islam M. [2010] Calcium signaling in the islets. *Adv Exp Med Biol* 654:235-59.
- [6] Duchen M, Verkhratsky A, Muallem S. [2008] Mitochondria and calcium in health and disease. *Cell Calcium* 44(1):1-5.
- [7] Zhu M, Ma J, Parrington J, Calcraft P, Galione A, Evans A. [2010] Calcium signaling via two-pore channels: local or global, that is the question. *Am J Physiol Cell Physiol* 298(3):C430-41.
- [8] Lipp P, Thomas D, Berridge M, Bootman M. [1997] Nuclear calcium signalling by individual cytoplasmic calcium puffs. *EMBO J* 16(23):7166-73.



- [9] Wu X, Zhang T, Bossuyt J, Li X, McKinsey T, Dedman J, et al. [2006] Local InsP3-dependent perinuclear Ca2+ signaling in cardiac myocyte excitation-transcription coupling. *J Clin Invest* 116(3):675-82.
- [10] Hardingham G, Chawla S, Johnson C, Bading H. [1997] Distinct functions of nuclear and cytoplasmic calcium in the control of gene expression. *Nature* 385(6613):260-5.
- [11] Berridge M, Lipp P, Bootman M. [2000] The versatility and universality of calcium signalling. *Nat Rev Mol Cell Biol* 1(1):11-21.
- [12] Yang F, Cui Y, Wang K, Zheng J. [2010] Thermosensitive TRP channel pore turret is part of the temperature activation pathway. *Proc Natl Acad Sci U S A* 107(15):7083-8.
- [13] Leung H, Tseng-Crank J, Kim E, Mahapatra C, Shino S, Zhou Y, et al. [2008] DAG lipase activity is necessary for TRP channel regulation in Drosophila photoreceptors. *Neuron* 58(6):884-96.
- [14] Inoue R, Jian Z, Kawarabayashi Y. [2009] Mechanosensitive TRP channels in cardiovascular pathophysiology. *Pharmacol Ther* 123(3):371-85.
- [15] Abramowitz J, Birnbaumer L. [2009] Physiology and pathophysiology of canonical transient receptor potential channels. *FASEB J* 23(2):297-328.
- [16] Sabatier N, Shibuya I, Dayanithi G. [2004] Intracellular calcium increase and somatodendritic vasopressin release by vasopressin receptor agonists in the rat supraoptic nucleus: involvement of multiple intracellular transduction signals. *J Neuroendocrinol* 16(3):221-36.
- [17] Wang S, Song L, Lakatta E, Cheng H. [2001] Ca2+ signalling between single L-type Ca2+ channels and ryanodine receptors in heart cells. *Nature* 410(6828):592-6.
- [18] Navedo M, Takeda Y, Nieves-Cintrón M, Molkentin J, Santana L. [2010] Elevated Ca2+ sparklet activity during acute hyperglycemia and diabetes in cerebral arterial smooth muscle cells. *Am J Physiol Cell Physiol* 298(2):C211-20.
- [19] Cheng H, Lederer W, Cannell M. [1993] Calcium sparks: elementary events underlying excitation-contraction coupling in heart muscle. *Science* 262(5134):740-4.
- [20] Niggli E, Shirokova N. [2007] A guide to sparkology: the taxonomy of elementary cellular Ca2+ signaling events. *Cell Calcium* 42(4-5):379-87.
- [21] Wray S, Burdyga T. [2010] Sarcoplasmic reticulum function in smooth muscle. *Physiol Rev* 90(1):113-78.
- [22] Lipp P, Niggli E. [1998] Fundamental calcium release events revealed by two-photon excitation photolysis of caged calcium in Guinea-pig cardiac myocytes. *J Physiol* 508 (Pt 3):801-9.
- [23] Niggli E. [1999] Localized intracellular calcium signaling in muscle: calcium sparks and calcium quarks. *Annu Rev Physiol* 61:311-35.
- [24] Taufiq-Ur-Rahman, Skupin A, Falcke M, Taylor C. [2009] Clustering of InsP3 receptors by InsP3 retunes their regulation by InsP3 and Ca2+. *Nature* 458(7238):655-9.
- [25] Parker I, Yao Y. [1996] Ca2+ transients associated with openings of inositol trisphosphate-gated channels in Xenopus oocytes. J Physiol 491 (Pt 3):663-8.

- [26] Koizumi S, Bootman M, Bobanović L, Schell M, Berridge M, Lipp P. [1999] Characterization of elementary Ca2+ release signals in NGF-differentiated PC12 cells and hippocampal neurons. *Neuron* 22(1):125-37.
- [27] Bayguinov O, Hagen B, Bonev A, Nelson M, Sanders K. [2000] Intracellular calcium events activated by ATP in murine colonic myocytes. *Am J Physiol Cell Physiol* 279(1):C126-35.
- [28] Pacher P, Thomas A, Hajnóczky G. [2002] Ca2+ marks: miniature calcium signals in single mitochondria driven by ryanodine receptors. *Proc Natl Acad Sci U S A* 99(4):2380-5.
- [29] Shannon T, Guo T, Bers D. [2003] Ca2+ scraps: local depletions of free [Ca2+] in cardiac sarcoplasmic reticulum during contractions leave substantial Ca2+ reserve. *Circ Res* 93(1):40-5.
- [30] Brochet D, Yang D, Di Maio A, Lederer W, Franzini-Armstrong C, Cheng H. [2005] Ca2+ blinks: rapid nanoscopic store calcium signaling. *Proc Natl Acad Sci U S A* 102(8):3099-104.
- [31] Wang S, Wei C, Zhao G, Brochet D, Shen J, Song L, et al. [2004] Imaging microdomain Ca2+ in muscle cells. *Circ Res* 94(8):1011-22.
- [32] Keller M, Kao J, Egger M, Niggli E. [2007] Calcium waves driven by "sensitization" wave-fronts. *Cardiovasc Res* 74(1):39-45.
- [33] Reither G, Schaefer M, Lipp P. [2006] PKCalpha: a versatile key for decoding the cellular calcium toolkit. *J Cell Biol* 174(4):521-33.
- [34] Carafoli E. [2004] The ambivalent nature of the calcium signal. *J Endocrinol Invest* 27(6 Suppl):134-6.
- [35] Toescu E, Möller T, Kettenmann H, Verkhratsky A. [1998] Long-term activation of capacitative Ca2+ entry in mouse microglial cells. *Neuroscience* 86(3):925-35.
- [36] Michelangeli F, Ogunbayo O, Wootton L. [2005] A plethora of interacting organellar Ca2+ stores. *Curr Opin Cell Biol* 17(2):135-40.
- [37] Sasaki N, Dayanithi G, Shibuya I. [2005] Ca2+ clearance mechanisms in neurohypophysial terminals of the rat. *Cell Calcium* 37(1):45-56.
- [38] Bano D, Nicotera P. [2007] Ca2+ signals and neuronal death in brain ischemia. *Stroke* 38(2 Suppl):674-6.
- [39] Toth A, Nickson P, Mandl A, Bannister M, Toth K, Erhardt P. [2007] Endoplasmic reticulum stress as a novel therapeutic target in heart diseases. *Cardiovasc Hematol Disord Drug Targets* 7(3):205-18.
- [40] Berridge M. [2010] Calcium hypothesis of Alzheimer's disease. *Pflugers Arch* 459(3):441-9.



ABOUT AUTHORS



Dr. Cedric Viero, Ph.D., is Research Associate in the Department of Cardiology, Wales Heart Research Institute, School of Medicine, Cardiff University, UK. Dr. Viero specializes in ion channel, calcium signalling and cardiac pathophysiology. He is a current member of the Working Group on Cardiac Cellular Electrophysiology (European Society of Cardiology). In his cardiovascular basic research career, he has authored more than 20 articles and abstracts for conferences, and contributed to 3 book chapters. Recently, he served as reviewer and editorial team member for peer-reviewed, international professional journals.



Prof. Govindan Dayanithi, Ph.D., is Research Director in CNRS-France. He was educated in India (University of Madras) as a Zoologist in 1983 and went to Europe on various fellowships/positions (Ministry of Education, Medical Res Foundation-France; Alexander von Humboldt Stiftung–Germany; JSPS-Japan; UMASS-USA; faculty of Medicine-Strasbourg; University of Oxford; employed by the French Ministry of Research in 1989. He is currently running a research group at the Institute of Experimental Medicine, Academy of Sciences of the Czech Republic, an EU Centre of Excellence in Prague. He serves as an Editorial Board member of the Journal of Neuroendocrinology, Journal of Endocrinology, Cell Calcium and International Journal of Cell Biology. His major research focus is to understand the role and physiology of calcium and calcium signalling in various tissue models and neuropeptide release. In collaboration with Japanese and British colleagues, he has employed newly generated genetically modified rat models for visualizing an AVP-enhanced green fluorescent protein (eGFP) fusion gene and an OT-enhanced cyan fluorescent protein (eCFP) fusion gene in order to understand the physiology of these neuropeptides in various tissue models. He is also studying the molecular physiology of calcium signalling in the human embryonic stem cells.